A Structural Model of the Type 1 Copper Protein Active Site: N₂S(thiolate)S(thioether) Ligation in a Cu(II) Complex

Patrick L. Holland and William B. Tolman*

Department of Chemistry and Center for Metals in Biocatalysis, University of Minnesota 207 Pleasant St. SE, Minneapolis, Minnesota 55455 Received April 17, 2000

The ubiquitous type 1 copper biological electron-transfer sites have a single Cu ion bonded to two histidine imidazoles and a cysteine thiolate (Cu–S \sim 2.15 Å) in an approximately trigonal geometry, usually with a fourth donor (typically a methionine thioether) present at a long yet bonding distance $(2.6-2.9 \text{ Å})^{-1}$ Recent work has shown that this classic motif is in fact variable, and there is great interest in understanding how the subtle changes in the Cu coordination sphere in the various type 1 sites relate to differences in their spectral and redox properties.^{1,2} Studies of synthetic copper complexes are helpful for understanding such structure/property relationships.³ Despite attempts over several decades to prepare inorganic complexes that reproduce the type 1 coordination geometry, however, no Cu(II) compound with sole N₂S(thiolate)S(thioether) ligation has been characterized definitively.4,5 We describe herein the successful attainment of this longstanding goal.

We recently reported⁶ the synthesis of the first three-coordinate Cu(II) complexes, including thiolate **1** that models the trigonal type 1 site in fungal laccase,⁷ by using a highly hindered β -diketiminate as a supporting ligand (L).⁸ In an extension of this approach, we treated solutions of LCuCl in THF with NaSC-(Ph)₂CH₂ECH₃ (E = O or S).⁹ For the case of E = O, product **2** was isolated as a stable blue-purple crystalline solid. The pendant ether is not coordinated to the Cu(II) ion in the complex and it adopts a solution geometry similar to that of **1**, as shown by



spectral data (Figures 1 and S1). Thus, the UV-vis and EPR spectral features of 1 and 2 are nearly identical, key features being the analogous $S \rightarrow Cu(II)$ LMCT band energies and intensities

(4) (a) Mandal, S.; Das, G.; Singh, R.; Shukla, R.; Bharadwaj, P. K. Coord. Chem. Rev. 1997, 160, 191. (b) Kitajima, N. Adv. Inorg. Chem. 1992, 39, 1.
(c) Bouwman, E.; Driessen, W. L.; Reedijk, J. Coord. Chem. Rev. 1990, 104, 143. (d) EPR and UV-vis spectra were obtained for a complex proposed to contain a N₂S(thiolate)S(thioether) donor set, but they do not resemble spectra of type 1 or 1.5 sites: Mandal, S.; Bharadwaj, P. K. Indian J. Chem. 1991, 30A, 948.

(5) Synthetic type 1 copper protein design efforts: Hellinga, H. W. J. Am. Chem. Soc. **1998**, *120*, 10055 and references therein.

(6) Holland, P. L.; Tolman, W. B. J. Am. Chem. Soc. 1999, 121, 7270.
(7) Ducros, V.; Brzozowski, A. M.; Wilson, K. S.; Brown, H.; Østergaard, P.; Schneider, P.; Yaver, D.; Pedersen, A. H.; Davies, G. J. Nat. Stuct. Biol 1998, 5, 310.



Figure 1. (a) X-band EPR spectra of toluene solutions of 2 (dashed line) and 3 (solid line), T = 20 K. (b) UV-visible spectra of solutions in heptane (1), pentane (2), or toluene (3) at ambient temperature.

as well as axial EPR signal parameters.¹⁰ The X-ray crystal structure of **2** (Figure 2a) also is similar to that of **1**. The Cu geometry is trigonal planar [the Cu atom lies 0.125(2) Å from the N₂S plane], and the metrical parameters mimic the coordination sphere of the three-coordinate type 1 Cu(II) site in fungal laccase in much the same way as described earlier for **1**.⁶

In contrast, using a ligand with a thioether functionality (E = S) in the place of the ether resulted in the formation of a product (3) with significantly perturbed spectroscopic and structural features. Importantly, the X-ray crystal structure of 3 (Figure 2b) reveals coordination of the thioether group to the metal ion [Cu-S(2) = 2.403(1) Å] to yield the first example of a structurally defined Cu(II) complex with the same donor set as the classical type 1 biological site. As a result of strong thioether binding, the remainder of the Cu coordination sphere in 3 differs significantly from those of 1 and 2. The metal–ligand distances in 3 are lengthened, consistent with its higher metal coordination number. In addition, in 3 the β -diketiminate ligand is "folded" along the

^{(1) (}a) Adman, E. T. Adv. Protein Chem. **1991**, 42, 145. (b) Solomon, E. I.; Baldwin, M. J.; Lowery, M. D. Chem. Rev. **1992**, 92, 521. (c) Randall, D. W.; Gamelin, D. R.; LaCroix, L. B.; Solomon, E. I. J. Biol. Inorg. Chem. **2000**, 5, 16.

⁽²⁾ Representative literature: (a) Palmer, A. E.; Randall, D. W.; Xu, F.; Solomon, E. I. J. Am. Chem. Soc. 1999, 121, 7138. (b) Buning, C.; Canters, G. W.; Comba, P.; Dennison, C.; Jeuken, L.; Melter, M.; Sanders-Loehr, J. J. Am. Chem. Soc. 2000, 122, 204. (c) Pierloot, K.; De Kerpel, J. O. A.; Ryde, U.; Olsson, M. H. M.; Roos, B. O. J. Am. Chem. Soc. 1998, 120, 13156. (d) LaCroix, L. B.; Randall, D. W.; Nersissian, A. M.; Hoitink, C. W. G.; Canters, G. W.; Valentine, J. S.; Solomon, E. I. J. Am. Chem. Soc. 1998, 120, 9621. (3) Karlin, K. D. Science 1993, 261, 701.

⁽⁸⁾ Feldman, J.; McLain, S. J.; Parthasarathy, A.; Marshall, W. J.; Calabrese, J. C.; Arthur, S. D. Organometallics **1997**, *16*, 1514.

⁽⁹⁾ For synthetic procedures and complete characterization data, see the Supporting Information.

Supporting information. (10) Data for **2** (yield 54%): UV/vis (pentane) [λ_{ex} , nm (ϵ , mM⁻¹ cm⁻¹)] 343 (~35), 423 (1.4), 480 (1.6), 566 (1.4), 738 (5.6); EPR (9.61 GHz, toluene, 20 K): $g_{II} = 2.17$, $A_{II}^{Cu} = 111 \times 10^{-4}$ cm⁻¹, $g_{\perp} = 2.04$, $A_{\perp}^{N} = 13 \times 10^{-4}$ cm⁻¹; cyclic voltammogram (0.2 M NBu₄PF₆/THF electrolyte, referenced to internal ferrocene): $E_{1/2} = -0.12$ V vs NHE, $E_{pa} - E_{pc} = 0.12$ V, $i_a \approx i_c$. Anal. Calcd for C4₄H₅₆CuN₂OS: C, 72.94; H, 7.79; N, 3.87. Found: C, 72.78; H, 7.89; N, 3.84. Data for **3** (yield 54%): UV/vis (toluene) [λ_{ex} , nm (ϵ , mM⁻¹ cm⁻¹)] 354 (~15), 430 (1.9), 538 (2.1), 691 (2.3); EPR (9.61 GHz, toluene, 20 K): $g_{II} = 2.15$, $A_{II}^{Cu} = 98 \times 10^{-4}$ cm⁻¹, $g_x = 2.01$, $g_y = 2.06$, $A_x^N = A_y^N$ = 10 × 10⁻⁴ cm⁻¹; cyclic voltammogram (0.2 M NBu₄PF₆/THF electrolyte, referenced to internal ferrocene): -0.20 V vs NHE, $E_{pa} - E_{pc} = 0.11$ V, $i_a \approx i_c$. Anal. Calcd for C4₄H₅₆CuN₂S₂: C, 71.36; H, 7.62; N, 3.78. Found: C, 71.92; H, 8.23; N, 3.48. Selected data for **1** (ref 6): $\lambda_{max} = 749$ nm, $\epsilon = 5.8$ mM⁻¹ cm⁻¹; $E_{I/2} = -0.18$ V vs NHE.



Figure 2. Thermal-ellipsoid plots (50% probability) of (a) **2** and (b) **3**, with hydrogen atoms omitted for clarity. Selected bond distances and angles: **2**, Cu-S = 2.119(1) Å; Cu-N = 1.900(4), 1.908(4) Å; $Cu-S-C = 112.3(1)^{\circ}$. **3**, Cu-S(1) = 2.242(1) Å, Cu-S(2) = 2.403(1) Å, Cu-N = 1.987(3), 1.952(3) Å, $Cu-S(1)-C = 109.4(1)^{\circ}$.

N-N vector [fold angle = $14.6(2)^{\circ}$] and the metal adopts a flattened tetrahedral geometry [55.10(8)° twist angle between the N_2Cu and S_2Cu planes] with the copper atom residing 0.484(2) Å above the N₂S(thiolate) plane (Figure S4). In contrast to the trigonal ligand disposition in 1 and 2 which results in very similar S(thiolate)-Cu-N angles (average 130.5°; range 126-136°), these angles in **3** differ considerably $[98.29(9) \text{ and } 146.53(9)^{\circ}]$. Because of this distortion, the overall geometry of 3 does not accurately replicate that of the "classic" type 1 sites (e.g. plastocyanin and azurin)¹ despite the identity of the donor atoms. The structure of 3 does, however, resemble the "Type 1.5" sites (e.g. nitrite reductase and several azurin mutants containing Gln, His, or Glu in place of the Met ligand)¹¹ that are characterized by stronger axial ligand interactions and a tetragonal distortion relative to the $\sim C_{3v}$ geometry of the type 1 centers.^{1c} This resemblance is illustrated by a comparison of the core of 3 to the relevant copper coordination spheres in the crystallographically determined structures of nitrite reductase and of M121H azurin (Figure 3).



Figure 3. Comparison of the coordination sphere of 3 to those of structurally characterized "perturbed" type 1 copper cores.

The spectral characteristics of 3 suggest that the differences from 2 seen in the solid state are retained in solution (Figure 1). In the EPR spectrum of **3**, the A_{\parallel}^{Cu} and g_{\parallel} values (98 \times 10⁻⁴ $\rm cm^{-1}$ and 2.15, respectively) are smaller than those for 2 (111 \times 10^{-4} cm⁻¹ and 2.17). In addition, the signal for **3** is rhombic: our best simulation has the g_{\perp} region split into $g_{r} = 2.01$ and g_{ν} = 2.06. The EPR spectrum is quite different from an independently prepared tetragonal copper(II) complex of L.12 The differences between the EPR data for 2 and 3 parallel those between the type 1 and 1.5 protein sites.¹³ Compared to 1 and 2, the electronic absorption spectrum of 3 shows a substantial shift of the characteristic low-energy $S \rightarrow Cu LMCT$ band to 691 nm, along with a decrease in its intensity ($\epsilon = 2300 \text{ M}^{-1} \text{ cm}^{-1}$). These data are also consistent with a four-coordinate solution structure, and show a striking resemblance to the changes observed upon deprotonation of Glu in the Met121Glu mutant of azurin.^{11b} These changes may be attributed to a longer Cu-S(thiolate) bond.^{13,14} Interestingly, cyclic voltammograms of 1-3 contain pseudoreversible waves with redox potentials that fall within a narrow range $(E_{1/2} - 0.12 \text{ to } -0.20 \text{ V vs NHE})$.¹⁰

In conclusion, by examining complexes that differ by only a single donor atom (E = O vs S), we have been able to address directly in a model system the structural and spectroscopic effects of thioether coordination to the type 1 N₂S(thiolate)Cu core. The thioether ligand in complex 3 is strongly bound, leading to a flattened tetrahedral structure distinct from the geometries of the three-coordinate molecules 1 and 2. This geometric difference is reflected in divergent spectral features. Structurally, complex 3 most closely matches the perturbed "type 1.5" biological sites, with similarities and differences in spectroscopic and redox properties that warrant further study.

Acknowledgment. We thank Profs. Lawrence Que, Jr. and John Lipscomb for providing access to resonance Raman/electrochemical and EPR equipment, respectively, and the NIH (GM47365) for financial support.

Supporting Information Available: Experimental details for all new compounds, EPR simulations, resonance Raman spectra, cyclic voltammograms (PDF) and X-ray crystallographic files (CIF) for **2** and **3**. This material is available free of charge via the Internet at http://pubs.acs.org.

JA001328V

^{(11) (}a) Romero, A.; Hoitink, C. W. G.; Nar, H.; Huber, R.; Messerschmidt, A.; Canters, G. W. J. Mol. Biol. **1993**, 229, 1007. (b) Karlsson, B. G.; Tsai, L.-C.; Nar, H.; Sanders-Loehr, J.; Bonander, N.; Langer, V.; Sjölin, L. Biochemistry **1997**, 36, 4089. (c) Messerschmidt, A.; Prade, L.; Kroes, S. J.; Sanders-Loehr, J.; Huber, R.; Canters, G. W. Proc. Natl. Acad. Sci. U.S.A. **1998**, 95, 3443. (d) Dodd, F. E.; Van Beeumen, J.; Eady, R. R.; Hasnain, S. S. J. Mol. Biol. **1998**, 282, 369.

⁽¹²⁾ Selected parameters for the axial EPR signal of LCu(η^2 -O₂CCH₃): $g_{\parallel} = 2.20$ and $A_{\parallel} = 187 \times 10^{-4}$ cm⁻¹. Bowen, L. L.; Holland, P. L.; Tolman, W. B. Unpublished results.

⁽¹³⁾ LaCroix, L. B.; Shadle, S. E.; Wang, Y.; Averill, B. A.; Hedman, B.; Hodgson, K. O.; Solomon, E. I. J. Am. Chem. Soc. **1996**, 118, 7755.

⁽¹⁴⁾ Both 2 and 3 show numerous bands in the region 250-430 cm⁻¹ in their resonance Raman spectra ($\lambda_{ex} = 632.8$ nm) that have not yet been assigned (see Supporting Information). For 1, there is substantial evidence that a band at 430 cm⁻¹ is due to copper–sulfur stretching: Randall, D. W.; DeBeer, S.; Holland, P. L.; Hedman, B.; Hodgson, K. O.; Tolman, W. B.; Solomon, E. I. Submitted for publication.